

Key Parameters of Measles Virus Production for Oncolytic Virotherapy

¹Weiss, K., ¹D. Salzig, ²M.D. Mühlebach,
³K. Cichutek, ⁴R. Pörtner and ^{1,5,6}P. Czermak

¹Institute of Bioprocess Engineering and Pharmaceutical Technology,
Mittelhessen University of Applied Sciences, Giessen, Germany

²Oncolytic Measles Viruses and Vectored Vaccines, Paul-Ehrlich-Institut, Langen, Germany

³Division of Medical Biotechnology, Paul-Ehrlich-Institut, Langen, Germany

⁴Hamburg University of Technology, Hamburg, Germany

⁵Department of Chemical Engineering, Kansas State University, Manhattan KS, USA

⁶Faculty of Biology and Chemistry, Justus-Liebig-University of Giessen, Germany

Received 2012-04-23; Revised 2012-05-15; Accepted 2012-05-29

ABSTRACT

Attenuated measles virus has revealed selective tumor cell killing and is currently tested in clinical trials for the therapy of cancer patients. The amount of infectious particles per dose needed for oncolytic therapy can be more than a million times higher compared to vaccination. This requires highly effective production processes which guarantee the measles virus quantities needed for its use in cancer therapy. Referring to measles virus production itself, several factors are influencing process parameters and subsequent virus yields. These factors are medium optimization, feeding of nutrients, an optimal multiplicity of infection, localization of produced virus particles, temperature sensitivity and time of harvest. This review summarizes the available data concerning measles virus production in cell culture and factors influencing the virus yield.

Keywords: Measles Virus (MV), Subacute Sclerosing Panencephalitis (SSPE), Major Complement Protein (MCP), Cytopathic Effect (CPE)

1. INTRODUCTION

Measles is a highly infectious disease of the respiratory and lymphoid systems caused by Measles Virus (MV). Beside the typical rash and Koplik spots, this disease is clinically characterized by high fever and a considerably weakened state of health due to transient, distinct immune suppression. The course of disease is often relatively mild, but sometimes life-threatening. Deadly complications such as e.g., severe pneumonia or Subacute Sclerosing Panencephalitis (SSPE) occur with a fatality rate of about 1 in 1,000 patients (Sabella, 2010). Especially the life-threatening direct ethiopathology of measles can be prevented by vaccination using life-attenuated vaccine strains of MV. These vaccines have been used in millions of doses up to now, probably evoking life-long immunity and revealing an excellent safety profile. Derivatives of these vaccine strains are used

in current clinical trials for their efficacy in oncolytic virotherapy (Msaouel *et al.*, 2011).

The only natural hosts for MV are humans or non-human primates. MV as a member of the order *Mononegavirales* is an envelope, single-stranded, negative-sense RNA virus from the family *Paramyxoviridae* (genus *Morbillivirus*) (Navaratnarajah *et al.*, 2009). The virus is found as pleomorphic particles with a size of 120-270 nm. The two glycoproteins hemagglutinin (H protein) and fusion protein (F protein) are embedded in the lipid bilayer constituting the viral envelope. The matrix protein (M protein) covers the inner side of the viral envelope and modulates the activity of the viral glycoproteins (Cathomen *et al.*, 1998). The viral RNA is encased in a ribonucleocapsid structure. The ribonucleocapsid consists of the nucleocapsid protein tightly wrapping the RNA genome and the associated viral polymerase (L) and viral Phosphoprotein (P), a co-factor of the polymerase (Fig. 1A).

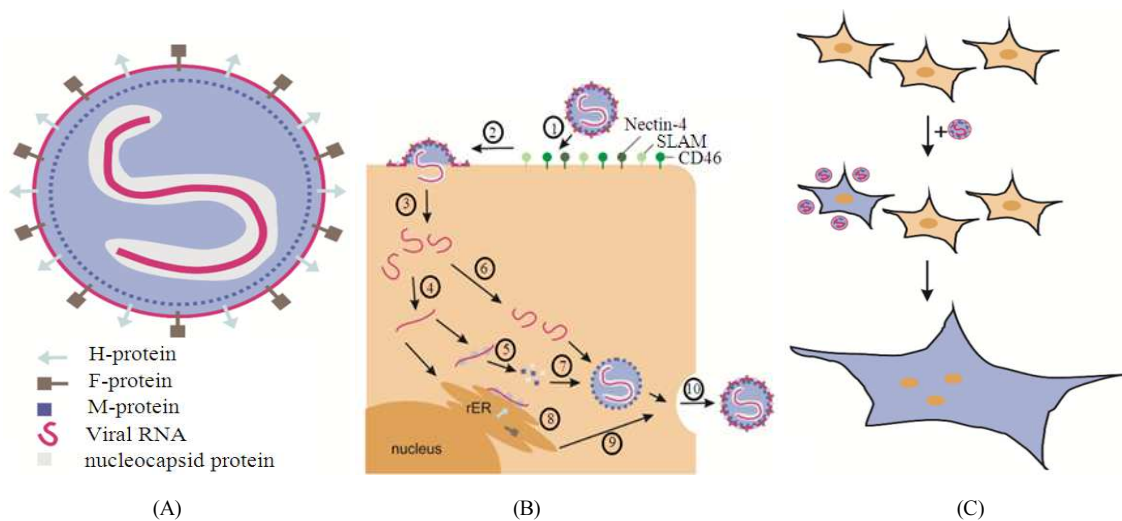


Fig. 1. Particle structure and replication cycle of MV. A: MV is a single-stranded RNA virus. Viral RNA is encapsulated by a viral nucleocapsid. The glycoproteins haemagglutinin (H protein) and fusion protein (F protein) are embedded in the lipid layer. The matrix protein (M-protein) covers the inner side of the viral envelope and interacts with viral glycoproteins and the ribonucleocapsid. B: MV attaches to the host cell membrane via the viral H protein binding to one of the three receptors SLAM, CD46, or nectin-4 (step 1). After receptor attachment, H is thought to trigger the associated F protein, resulting in fusion of viral and host cell membranes (step 2) with the formation of a fusion pore, through which the ribonucleoprotein complex enters the cells' cytoplasm (step 3). Viral RNA polymerase uses ribonucleotide triphosphates in the cytosol to transcribe the viral genome into viral mRNAs (step 4) and, after translation of viral proteins (step 5), subsequently to replicate the viral RNA genome and antigenome (step 6). Together with the replicated viral genome, the N, P and L proteins assemble a progeny ribonucleocapsid (step 7). H protein and F protein, the viral transmembrane glycoproteins, are produced at the ribosomes of the rough ER (step 8). These two proteins are transported as oligomeric complexes to the cell membrane via the Golgi apparatus utilizing the exocytotic pathway (step 9). The progeny ribonucleocapsid associates at the plasma membrane of the host cell with the viral transmembrane glycoproteins, mediated by the M protein. Direct association of the M protein with the cytoplasmic domains of the H and F proteins is supposed to induce folding of the cell membrane, followed by the release of the virus particle by budding (step 10). C: Infected cells fuse with neighboring receptor-positive cells resulting in multi-nucleated giant cells (syncytia).

The attachment protein of MV, the H protein, exhibits no neuraminidase activity, as already indicated by its name, nor does it adsorb to neuraminic acid as entry receptor, which is otherwise typical for paramyxoviral attachment proteins of other genera (Navaratnarajah *et al.*, 2009). The H and the F proteins mediate MV cell entry at the cell surface via fusion of the viral lipid envelope bilayer and the host cell cytoplasmic membrane (**Fig. 1B**). The H protein binds to specific host cell receptors and is thought to trigger the F protein via conformational changes to initiate the fusion between the virus' and the cells' membranes. Three main receptors are used by MV. Pathogenic wild-type strains of MV utilize signaling lymphocyte activation molecule (SLAM/CD150) (Tatsuo *et al.*, 2000) on activated immune cells and nectin-4/PVRL-4 (Mühlebach *et al.*, 2011; Noyce *et al.*, 2011) on epithelial cells as receptors. The vaccine strain has expanded its receptor specificity and additionally uses CD46, i.e., Major Complement Protein (MCP) (Dorig *et al.*, 1993; Gerlier *et al.*, 1994), which is

expressed on all human nucleated cells, for entry (Navaratnarajah *et al.*, 2009). After infection, MV induces morphological changes of the cell layer, a so called Cytopathic Effect (CPE) in infected cell cultures. MV infection is accompanied by cell-to-cell fusion and formation of multi-nucleated giant cells, so called syncytia (**Fig. 1C**). In the late stages of MV infection, apoptosis and the lysis of the cells are induced. During the amplification phase, a considerable part of the MV particles remains cell-associated, while some particles are released from the cell membrane via budding (Morgan and Rapp, 1977).

1.1. Application of Attenuated Measles Virus

The initial isolation of the MV was done by Enders and Peebles (1954). The isolated strain has been derived from the throat washings of an 11 years old measles patient named David Edmonston and was therefore named Edmonston strain (Enders and Peebles, 1954). This measles virus strain was adapted to cell culture and

attenuated by serial tissue culture passages resulting in less virulent virus strains, e.g., Edmonston B. Thus, this isolate has become the progenitor for many attenuated strains which are currently in the use for vaccination, (Fig. 2A) or research investigation.

1.2. Use of Measles Virus For Vaccination

It was Thomas Peebles who obtained specimens from a measles outbreak in 1954. Meanwhile, human kidney cells had been successfully cultured *in vitro* in the Enders lab.

In these cells, MV could be successfully cultivated for the first time and has been subsequently passaged 23 times. After inoculation of these passaged virus particles in measles-susceptible monkeys (cynomolgus monkeys), the animals developed fever, rash, viremia and anti-MV antibodies (Peebles *et al.*, 1957). After one further passage, the cultured MV had been transferred to human amnion cell cultures and then to chicken eggs for 28 and 6 passages, respectively (Milovanovic *et al.*, 1957). Afterwards, the virus was passaged in chicken embryo fibroblasts for 13 times. When the virus particles were then inoculated into measles-susceptible monkeys, no disease symptoms, but complement-fixing and neutralizing-antibodies were observed (Katz *et al.*, 1958; Enders *et al.*, 1960). These monkeys were then resistant to infection with “wild type”, pathogenic MV. Based on these results a vaccine strain was developed and licensed in 1963 (Katz, 2009). Thus, measles vaccination became possible that enables control of measles epidemics.

Today, several MV strains are used and licensed for vaccination, e.g., the Moraten Vaccine (Merck) (Scheld *et al.*, 2004) or AIK-C, Schwarz F88, CAM-70 and TD97, (Fig. 2B) (Artenstein, 2009). These vaccines have been used in millions of doses and have revealed an excellent safety profile.

Measles vaccine is currently produced by standard cell culture techniques in chicken embryo fibroblasts (Bronzino, 2000). The used process systems are mainly roller bottles or multilayer stacked plate systems (cell factories). Compared to petri dishes or T-flasks, a larger surface can be obtained with these systems.

1.3. Potential of Measles Virus for Cancer Therapy

Apart from vaccination, attenuated MV has a natural ability to specifically kill cancer cells. Attenuated MV enters the cell via the receptor CD46, which is frequently over-expressed on tumor cells (Davis and Fang, 2005). As MV is a lytic virus, MV replication in tumor cells leads to cell lysis of cancer cells. Intratumoral injection of the unmodified MV Edmonston strain or an Edmonston strain derived recombinant MV induced

regression of large established human lymphoma xenografts in murine models (Grote *et al.*, 2001). This oncolytic virus has also been reported to have potency against human epithelial ovarian cancer cells *in vitro* and *in vivo*, being selectively lytic for the ovarian tumor cells (Peng *et al.*, 2002), hepatocellular carcinoma (Blechacz *et al.*, 2006), or prostate cancer cells (Msaouel *et al.*, 2009b), among others. Russell *et al.* (2010) filed a patent application which describes a method for limiting the growth of cancer cells using an attenuated MV strain. This oncolytic property qualifies MV for cancer therapy approaches and is currently examined in clinical trials with human cancer patients (Davis and Fang, 2005). Furthermore, new oncolytic MVs have been created as next generation therapeutics, which specifically target the cancer cells (Nakamura *et al.*, 2005; Springfield *et al.*, 2006; Hasegawa *et al.*, 2007; Mühlebach *et al.*, 2010; Leber *et al.*, 2011).

One problem of using oncolytic measles viruses for cancer treatment is the sheer amount of virus needed. Similar to virus particle based gene therapy (Nehring *et al.*, 2004; 2006; 2009) the measles virus quantity for a therapeutic dose is three to 6 log₁₀ units higher than for vaccination (Msaouel *et al.*, 2009a; 2011; Davis and Fang, 2005; Russell *et al.*, 2010). Searching for a production process for oncolytic virus particles the question arises, whether the production process for MV vaccines cannot simply be expanded. To answer this question, the specific requirements for MV production are discussed in this review.

1.4. Oncolytic MV Particles: New Aspects for MV Production

Currently, there is just one detailed description for MV production in larger scales (60 L) for oncolytic therapy available, where a detailed protocol for the purification and the serum-free production with Vero cells in cell factories is presented (Langfield *et al.*, 2011). Measles vaccines are routinely still produced using chicken embryo fibroblasts and standard cell culture techniques. While this process works for that specific purpose, it is not easily transferable for oncolytic virotherapy applications due to the up to 6 log₁₀ units higher amounts of virus needed. Therefore, a massive scale up of the production process is required. All currently industrial used processes for measles vaccine production are monolayer cultures with non-uniform cell growth, limiting oxygen transport and inhomogeneous nutrient supply. From the bioprocess point of view, these processes are neither controllable, nor do they yield standardized and reproducible virus qualities and quantities.

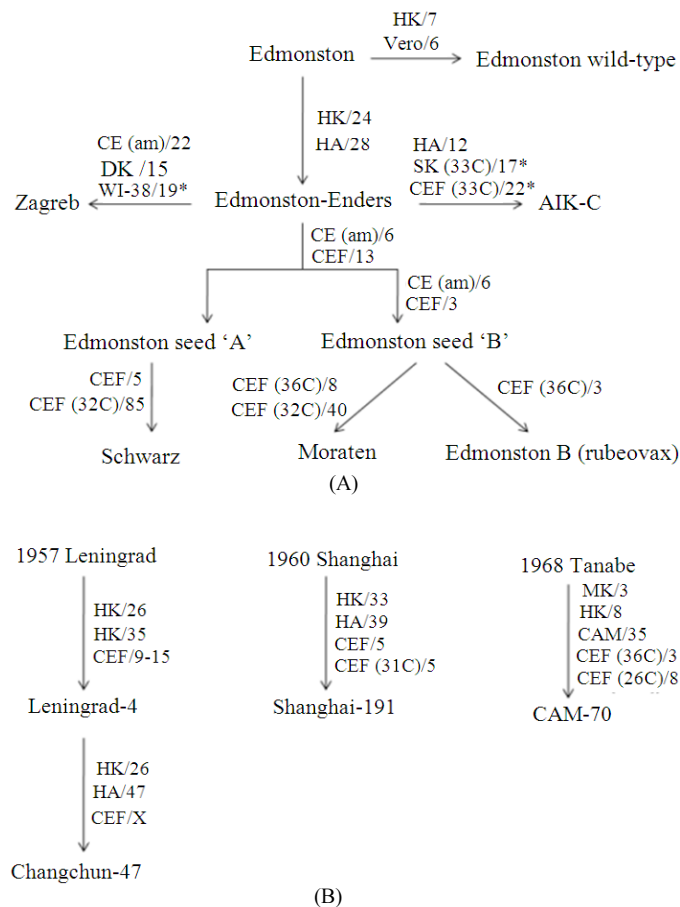


Fig. 2. Histories of Edmonston derived (A) and non-Edmonston (B) derived vaccines (obtained from (Bankamp *et al.*, 2011), with permission). Derived strains are indicated by the abbreviation of the cell culture used and the number of passages performed. The temperature was assumed to be 37°C, other used temperatures were indicated by numbers in brackets. CAM...chorioallantoic cavity of chick embryo; CE(am)...intraamniotic cavity of chick embryo; CEF...chick embryo fibroblast; DK...dog kidney; HA...human amnion; HK...human kidney; SK...sheep kidney; WI-38...human diploid cells; *Plaque purification.

Furthermore, an efficient scale up of these processes is difficult and requires expensive robotic automations (Bronzino, 2000; Trabelsi *et al.*, 2010). To expand these existing systems in a way that over 1 million times higher numbers of infectious particles per dose are produced, would be, even if possible, very inefficient.

Another point to consider is the virus per se, especially for those oncolytic virus particles that are fully retargeted for cancer cells. For the latter, the virus recognizes only receptors expressed on human cancer cells and propagation of these oncolytic viruses in chicken embryo cells will therefore not be possible. Thus, at least some oncolytic MV viruses have to be produced in special cells developed for this purpose, e.g. Vero-HIS cells

(Nakamura *et al.*, 2005). For these special cells, a new process has to be established, anyway. Fermentation techniques enable high efficient production of very large volumes to provide the necessary quantity of the desired product. Production processes in bioreactors reduce overall costs and shortens time. This can easily be presented by evaluating the productivity (space-time yield), which describes the ratio of the amount of desired product, that is generated and the time and volume needed for this purpose. The productivity of conventional cultivation systems (e.g., cell factories) compared to bioreactors is supposed to be lower.

Scaling up in a bioreactor requires an increase in the reactor vessel while the operating effort stays the same.

In contrast a scale up in a cell factory production process means increasing the number of cell factories. These additional cell factories have to be handled, which increases required material, staff costs and handling time.

Besides, bioreactor systems are characterized by its complex design and high level of instrumentation which enables online measurement and in process control with reproducibility efforts and GMP standardization. In **Table 1** the main advantages and disadvantages of the two systems are compared.

The purpose of this review is to summarize published production processes for MV and factors influencing the virus yield. Moreover, it indicates starting points for further analysis to obtain sufficient amount of virus particles for oncolytic therapy. Other aspects of MV virotherapy have been reviewed already and for this reason were not covered in this review. Details covering MV biology are summarized in (Griffin and Oldstone, 2008), the current state of oncolytic trials with measles viruses is described by (Msaouel *et al.*, 2009a, Msaouel *et al.*, 2011). Additionally, Langfield *et al.* (2011) and Udem (1984) provide detailed description of methods for MV purification.

1.5. Measles Virus Production

Starting with the demonstration that MV could be propagated in human and simian cell cultures (Enders and Peebles, 1954), a considerable amount of data has been published on MV production. MV amplification has been described in various cell lines and with various culture modes. Parameters such as virus stability, composition of medium and virus amount have already been analyzed. Concerning an optimization of the production process, only few data have been published. The outcome in the published data was evaluated by the cell-associated and extracellular virus titers. For MV, two main methods evaluating titers of infectious MV are used. For infectivity

titration, virus suspensions are diluted and endpoint dilutions are determined, at which infection is detectable (often by the appearance of syncytia) (Reed and Muench, 1938). Using statistics, virus titers are subsequently calculated as Tissue Culture Infective Dose (TCID₅₀) (Kärber, 1931), which indicates the dilution of the respective virus particle suspension, at which infection is detectable in 50% of inoculated cultures. Another option of virus quantification is the evaluation of plaque formation. Employing this method, cells are infected with various virus dilutions and after an adsorption period of about two hours, a soft agar overlay containing all medium compositions is added (Mayer, 1960; Rapp, 1964). Due to the spread of the virus via cell lysis, areas of virus-induced cell destruction appear as so called plaques. The number of plaques is then counted after a defined period of time. According to this method the virus titers are given in plaque forming units (pfu). Thus, the pfu method describes an infectivity that is directly quantified, whereas the TCID₅₀ is a statistical description of a probability of infection with 50% chance. Accordingly, a titer of 1×10^5 TCID₅₀ mL⁻¹ corresponds to approximately 0.7×10^5 pfu mL⁻¹.

In the following, measles virus has been produced using serum supplemented medium, if not specified otherwise.

1.6. Impact of Virus Strain and Cell Type for Measles Virus Production

MV production has been analysed in several cell types and with several virus strains. Matumoto (1966) gives an overview of various cell lines, which had been analysed and could be useful for MV production. While research groups have determined various cell types for their susceptibility to MV, industrial MV production (vaccines) are still routinely produced using chicken embryo cells (Bronzino, 2000).

Table 1. Comparison of conventional (e.g., cell factories) and bioreactor system for cell culture cultivation and production. Presentation of the main advantages and disadvantages of the respective system

Conventional cultivation (e.g., cell factories)	bioreactor cultivation
Simple design	Often more complex design
Low level of instrumentation, but requires transport of the whole culture to e.g., work bench for medium exchange	Requires external equipment (e.g., heating circuit, sterile couplers)
Low level of control (limitation of metabolites)	Possibility of direct regulation for appropriate physical and chemical requirements
Due to static cultivation development of local concentration gradients	Increased homogeneity
Due to increased inhomogeneity reduced stability and reproducibility	Increased stability and reproducibility
Ease of handling in small scale	Easier to handle in larger scales
During cultivation measurement of parameters hard to perform	Online monitoring of process parameters with integrated sensor
Insufficient oxygen supply for high cell densities	Increased oxygen transfer into the medium

1.7. The Edmonston Strain

As already mentioned, the Edmonston strain is up to now the most commonly used strain for vaccine development. The maximum virus yield obtained with the Edmonston strain has been determined by different groups, while various cell lines were infected with this MV strain (Black, 1959; Kohn and Yassky, 1962; Nakai *et al.*, 1969; Romano and Scarlata, 1973; Bacsko and Lazzarini, 1979; Brandriss *et al.*, 1982; Scott and Choppin, 1982; Mendonca *et al.*, 1994; 2002) (**Table 2**).

Black (1959) gained a maximum titer of 6.3×10^5 pfu mL⁻¹ with Hep-2 cells, whereas Romano and Scarlata (1973) described a maximum titer of 4.6×10^4 TCID₅₀ mL⁻¹ in HeLa cells and Udem (1984) a maximum of 1×10^8 pfu/mL in HeLa cells. In KB cells, a maximum titer of 5.6×10^6 TCID₅₀ mL⁻¹ was obtained (Kohn and Yassky, 1962). A maximum value of 1×10^7 pfu mL⁻¹ was produced with BSC-1 cells (Nakai *et al.*, 1969), whereas MV produced in Vero cells had a maximum titer of 1.7×10^8 TCID₅₀ mL⁻¹ (Mendonca *et al.*, 1994; 2002). MV production in the suspension cell line U937 resulted in a titer of 6×10^6 pfu mL⁻¹ (Brandriss *et al.*, 1982). A comparison of Vero, BGMK, CV-1, Hep-2, AV₃, BHK-21, PK-15 and MDCK cells infected with the Edmonston strain revealed that Vero and BGMK cells produced the highest virus yields (Scott and Choppin, 1982). The yield with Vero cells was slightly higher with a maximum value of 2.8×10^8 pfu mL⁻¹. Other cell lines had reduced virus production, with 7-28,000 times lower titres. MDCK cells had the lowest production rate with just 1×10^4 pfu mL⁻¹ (Scott and Choppin, 1982).

A further analysis of MV production in Vero and MA160 cell monolayers as well as MA160 cells cultured in suspension has been published. A maximum titer of approx. 30 pfu per cell was described on MA160 monolayers, while the titers in Vero cells were approximately three times lower. In contrast, with suspension culture of MA160 cells, a yield of 100-800 pfu per cell (equals 2×10^7 - 1.6×10^8 pfu mL⁻¹) could be reached; this is an increase of 20-100 times compared to the monolayer titers. However, besides the parameters cell types and culture mode, the medium supplements have also been varied during the three culture set-ups (Bacsko and Lazzarini, 1979). Thus, no absolute comparison is feasible, since the impact of medium composition on MV production yields has already been documented, as well (Romano and Scarlata, 1973; Scott and Choppin, 1982). Taken together, these data may be helpful for choosing an optimal MV producer cell line, but have to be interpreted with respect to the composition of the medium.

1.8. Other Measles Virus Strains

Besides the Edmonston lineage, several other strains of MV have also been examined. These are the Hallé strain (Dubois-Dalcq and Reese, 1975; Scott and Choppin, 1982), the TYCSA strain (a variant of the Toyoshima strain) (Shishido *et al.*, 1967), the Leningrad-16 strain (Boriskin *et al.*, 1988; Sidorenko *et al.*, 1989) and the AIK-C strain (Trabelsi *et al.*, 2010) (**Table 2**).

Production of the Hallé strain in Vero cells was characterized by electron microscopy. Due to these data it has been suggested that infected Vero cells do not complete the first cycle of viral replication within 24 h after infection. At this time point, cell fusion was just starting; neither budding nor complete virus particles could be observed. In contrast, the whole monolayer appeared to be covered with syncytia after three days and viral budding could be illustrated (Dubois-Dalcq and Reese, 1975). These observations are in agreement with previous data published for MV growth curves for the Edmonston strain in other cell lines (Black, 1959; Nakai *et al.*, 1969).

Scott and Choppin (1982) compared the virus production between the Edmonston and the Hallé strains. Infection with the Edmonston strain resulted in slightly higher maximum MV yields compared to Hallé (approximately 1×10^8 vs. 2.5×10^8 pfu mL⁻¹, respectively).

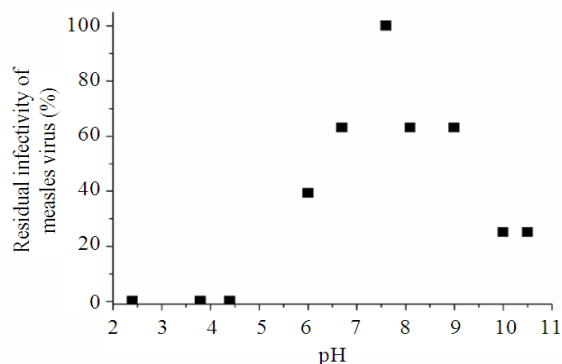
Vero cells infected with the TYCSA strain produced a maximum titer of 3×10^6 TCID₅₀ mL⁻¹ (Shishido *et al.*, 1967), whereas the Leningrad-16 strain amplified in quail embryo cells (Boriskin *et al.*, 1988; Sidorenko *et al.*, 1989), resulted in maximum titers of 1×10^7 or 3.6×10^5 TCID₅₀ mL⁻¹ (Boriskin *et al.*, 1988). Finally, infection of MRC-5 cells with the AIK-C strain resulted in a maximum titer of 1×10^{11} TCID₅₀ mL⁻¹ (Trabelsi *et al.*, 2010). This is to our knowledge the highest titer, which has been reported for MV, yet. **Table 2** summarizes maximum titers as discussed above. Direct comparison of various cell lines and measles virus strains has only been performed by (Scott and Choppin, 1982), so far, who documented the highest yields using Vero cells and the Edmonston strain. As other publications though reported higher yields with other cells and strains, the comparability of the data from different publications is not necessarily given. Especially for individual uses such as recombinant retargeted viruses for cancer therapy, investigations have to be performed with this strain anyway.

1.9. Stability of Measles Virus

Several parameters can influence the stability of MV particles. While some factors such as temperature or pH may inactivate the virus, others like certain salt supplements are able to stabilize the infectivity of the virus.

Table 2. Cell type and MV strain for MV production. Summary of the main virus strains which were produced with different cell lines. For better comparability, titers in pfu were converted in TCID₅₀ by multiplication with the factor 0.7

MV strain	Cell line	max. titer ($\times 10^6$ TCID ₅₀ /mL)	Ref.
Edmonston	AV ₃	2.800	(Scott and Choppin, 1982)
	BGMK	154.000	(Scott and Choppin, 1982)
	BHK-21	0.014	(Scott and Choppin, 1982)
	BSC-1	14.000	(Nakai <i>et al.</i> , 1969)
	CV-1	28.000	(Scott and Choppin, 1982)
	HeLa	0.046	(Romano and Scarlata, 1973)
		70.000	(Udem, 1984)
	Hep-2	8.400	(Scott and Choppin, 1982)
		0.630	(Black, 1959)
	KB	0.560	(Kohn and Yassky, 1962)
	MA160	230.000	(Baczko and Lazzarini, 1979)
	MDCK	0.007	(Scott and Choppin, 1982)
	PK-15	0.007	(Scott and Choppin, 1982)
	U937	8.600	(Brandriss <i>et al.</i> , 1982)
	Vero	196.000	(Scott and Choppin, 1982)
AIK-C	MRC-5	170.000	(Mendonca <i>et al.</i> , 1994)
		8.000	(Mendonca <i>et al.</i> , 2002)
		100,000.000	(Trabelsi <i>et al.</i> , 2010)
Leningrad-16 strain	quail embryo cells	10.000	(Trabelsi <i>et al.</i> , 2012)
		10.000	(Boriskin <i>et al.</i> , 1988)
		0.360	(Sidorenko <i>et al.</i> , 1989)
TYCSA	Vero	3.000	(Shishido <i>et al.</i> , 1967)
Halle		70.000	(Scott and Choppin, 1982)

**Fig. 3.** Residual infectivity of measles virus at various pH. MV was mixed with two volumes of cold buffer solutions of various pH values and incubated for 3 h in an ice bath. Determination of the virus titer was evaluated by titration (plaque method). Infectivity was normalized to optimal pH 7.6 and residual infectivity is plotted against pH. Figure is based on data published by Black (1959).

Temperature dependent inactivation of MV at 37°C has been reported to proceed at a rate of about 0.15 log₁₀ units per hour, which represents a viral half-life of about two hours (Black, 1959). Kohn and Yassky (1962) also reported a high inactivation rate of MV at 37°C, with a drop in infectivity by 90% in 16 h or 1.4 log₁₀ units per 24 h. Due to MV's high susceptibility

to temperature-inactivation (Black, 1959; Kohn and Yassky, 1962), there has been considerable interest in stabilizing the virus. Rapp *et al.* (1965) reported the impact of various salt solutions on the stability of MV. Dilutions of infectious particle preparations in 1 M MgSO₄ and 1 M Na₂SO₄ stabilized the virus. In contrast, in other salt solutions (CaCl₂, NaCl, KCl, K₂SO₄ and Na₂HPO₄), or in pure water, a decrease of one to more than three log₁₀ units was observed within 15 min at 50°C. With MgSO₄ and Na₂SO₄, only slight decreases of 0.2 or 0.4 log₁₀ units, respectively, were observed. Furthermore, MV inactivation at 50°C decreased with increasing MgSO₄ concentration. However, an increasing concentration of MgCl₂ enhanced the inactivation of MV. These results indicate that sulfate ions may protect MV against thermal inactivation (Rapp *et al.*, 1965).

Boriskin *et al.* (1988) analyzed the impact of MgSO₄ on MV yields, again (Boriskin *et al.*, 1988). Here, the same thermal inactivation rate of MV in the absence and presence of MgSO₄ was observed. It has been suggested that sulfate ions cannot protect the virus from heat inactivation in contrast to the previously cited publication (Rapp *et al.*, 1965). As an explanation for this discrepancy, the addition of MgSO₄ to the culture medium was described to result in earlier plaque formation and thus in an increase of apparent titers by up to 200-fold. MgCl₂ supplementation showed no stimulating effect and the addition of Na₂SO₄ showed a

maximum virus increase of apparent titer of only 10-fold (Boriskin *et al.*, 1988). Thus, certain ions may increase virus production or infectivity, but not its stability. This was further documented by analyzing MV NP protein synthesis by radioimmune precipitation and SDS-PAGE, where MV ribonucleocapsid protein appears in untreated cells to a much lesser extent, as compared to MgSO_4 treated cells (Boriskin *et al.*, 1988).

The pH sensitivity of MV has also been analyzed. **Figure 3** presents residual infectivity of MV at various pH (Black, 1959) showing a highest stability at pH 7.6 and a progressive inactivation of MV for both acidification and alkalization results.

MV particles are highly susceptible to temperature and pH dependent inactivation (Black, 1959; Kohn and Yassky, 1962; Rapp *et al.*, 1965; Boriskin *et al.*, 1988). Therefore it can be concluded, that during production released virus is inactivated very fast in the supernatant. For maximum virus yields, immediate cooling after the harvest, repeated harvesting in at least hourly intervals, further additional analyses for stabilizing the virus particles and online measurements and regulation of the pH during MV production are thus required.

1.10. Extracellular and Cell-Associated Virus

While production of MV components is inside the infected cell, all parts accumulate at the cell membrane and are released via budding. These active and complete MV particles appear extracellular. Other particles stay cell-associated. In the last phase of virus replication infectious virus as well as non-infectious virus components and particles can be released through cell disruption.

The derivation of the virus preparation is important, since extracellular virus is better to handle, as it is easy to separate from cells. Intracellular virus particles require cell disruption, thereby releasing potential impurities, i.e., cellular and viral components besides infectious particles, into the preparation.

One of the first reports about MV production in Hep-2 cells examined extracellular and cell-associated MV concentration during infection (Black, 1959). Although the cell disruption method, a one-step freeze-thaw cycle at -5°C , was referred to as incomplete, the amounts of extracellular virus were always lower than cell-associated yields. Infectious cell-associated virus had been found first after 15-18 h p.i. and extracellular virus could be observed about 26 h post infection. For both, maximum virus titers appeared after 48 h p.i. While cell-associated virus titers decreased after 48 h p.i. the yields for extracellular virus remained constant suggesting continuous virus release. This was confirmed by thermal-stability studies, as well. As previously mentioned, MV has a half-life of 2 h at 37°C (Black, 1959). Extracellular

virus titer could remain constant, because there seems to be equilibrium between virus release and decay.

In another study by Nakai *et al.* (1969) the yields of released virus and cell-associated virus were always within the same range (maximum titers of approximately 5×10^6 pfu mL^{-1}). Higher extracellular yields appeared with increasing cell-associated virus titers, also suggesting a continuous budding of virus particles (Nakai *et al.*, 1969). Interestingly, HeLa cells showed temperature dependant virus release. While at 37°C between a maximum of ~80 and 98% of the virus particles remained cell-associated, at 32.5°C more than 50% of the virus had been released not later than 60 h p.i. (Udem, 1984).

Shishido *et al.* (1967) infected Vero cells with a variant of the Toyoshima strain; MV titers started to increase for cell-associated virus about 12 h after infection, for extracellular virus about 18 h p.i. Titers decreased after 72 h p.i. for both, extracellular and cell-associated, virus populations. Maximum yields were about 3×10^6 for cell-associated and 5×10^5 TCID₅₀ mL^{-1} for released virus. Except for the end of the cultivation period, cell-associated virus titers were always one to two $\log_{10}$ units higher than extracellular titers (Shishido *et al.*, 1967). Accordingly, released virus in Vero and MA160 cells infected with the Edmonston strain represented only about 10% of the total amount of infectious virus (Baczko and Lazzarini, 1979). However, concentrations of cell-associated and extracellular virus were similar for BSC-1 cells infected with a virulent Edmonston derivative of MV, whereas attenuated virus remained to more than 95% cell-associated (Rapp, 1964).

These experiments were all performed in static systems, e.g., petri dishes. In bioreactor cultivation with Vero cells, the extracellular yield for MV of the Edmonston strain was lower than intracellular virus titer, as well. While the extracellular TCID₅₀ per mL was reduced by the value of about $1.1 \log_{10}$, the total virus titer was only lowered by $0.4 \log_{10}$ (Mendonca *et al.*, 1994).

Using the suspension cell line U937 in spinner system, less than 20% of the total amount appeared extracellular in spinner flasks (Brandriss *et al.*, 1982), whereas during production with the suspension cell line MA160 in spinner flasks about 10% of the total virus had been released (Baczko and Lazzarini, 1979).

Finally, the titers of cell-associated virus have almost always been reported to be higher than extracellular titers. Yields of cell-associated virus were mainly within a range of 50% up to 98% of total virus titers. Nevertheless, as already mentioned, serial harvesting of released virus should be considered. Since cell-associated virus can be harvested only once, cumulative titers of extracellular virus could exceed cell-associated yields.

1.11. Impact of Medium Composition on Measles Virus Production

In general, the medium composition has a great impact on the outcome of a production process. There are several components and supplements necessary or advantageous for cell cultivation and therefore virus production. Since the special requirements of infected cells are unknown, there is considerable need for detailed studies concerning these factors.

1.12. Impact of Serum

Serum, mostly Fetal Calf Serum (FCS), is generally supplemented to cell culture media as it provides growth factors, vitamins, hormones, amino acids, substances for adhesion and cytokines. It is supplemented for cell culture usually in amounts varying between 3 and 25% (v/v). Serum as a preparation of primary material is not completely characterized, but often essential. The use of serum-free fully defined media is often preferred for more robust and reproducible processes especially for the production of therapeutics.

The impact of serum content on MV production was analyzed by producing the Edmonston strain of MV on Vero cells in the absence and presence of 10% FCS. Titers (pfu per mL) were 0.5-1.5 log₁₀ units higher with 10% FCS than without (48 pfu per cell). With increasing FCS concentration (no serum, 2% and 10%) the virus yield increased. Substitution with calf or horse serum resulted in similar effects, although the use of FCS resulted in slightly higher titers. The addition of 10 mM HEPES to FCS (10%) supplemented medium resulted in a titer of 185 pfu per cell. Cultivation of infected cells without FCS, but with just 10 mM HEPES or just 5 µg mL⁻¹ insulin resulted in a virus titer of 65 or 123 pfu per cell, respectively, while the highest amount (191 pfu per cell) was obtained using all supplements (10% FCS, HEPES and insulin) (Scott and Choppin, 1982).

Using M199 Medium after infection with different supplements (none, 2% FCS, 0.2% gelatin, or 0.5% BSA) resulted in quite similar titers. The highest titer had been reached with BSA supplemented medium (10⁷ TCID₅₀ mL⁻¹). The lowest titer was equal to 3.2×10⁶ TCID₅₀ mL⁻¹ and resulted with M199 medium without any supplements (Trabelsi *et al.*, 2012).

1.13. Impact of Amino Acids in the Culture Medium

Most cell lines depend on supplemented amino acids to maintain their metabolism during *in vitro* cultivation. Two media have been compared with respect to microscopically detectable morphological changes of infected Hep-2 cells (Reissig *et al.*, 1956).

Enders medium contained bovine amniotic fluid, beef embryo extract, heated horse serum and Hanks balanced salt solution. Eagles (1955) medium was supplemented with amino acids and serous enrichments as described and has been additionally supplemented with calf serum. Up to tenfold higher virus yields were observed in Eagles medium compared to Enders medium (exact values had not been published). Additionally, the morphology of the cellular monolayer was different. In Enders medium, the monolayer was transformed into few large syncytia. In Eagles medium, the cells appeared more as a network and became rounded in shape (Fig. 4) (Reissig *et al.*, 1956).

When Enders medium was additionally supplemented with glutamine, the morphology of the cell sheet was similar to that in Eagles medium. The other way around, using glutamine deficient Eagle's medium, the cell morphology corresponds to the morphology of the cells in the normal Enders medium. This result suggests that the morphological changes were due to glutamine omission. Hence, the authors concluded, that the presence or absence of glutamine in the medium was the reason for the different virus titer (Reissig *et al.*, 1956).

The amino acid requirements for MV production have been examined in HeLa cells, as well (Romano and Scarlata, 1973). For that purpose, the omission or addition of certain amino acids (tryptophan, cysteine, tyrosine, leucine, valine, phenylalanine, isoleucine, threonine, methionine, lysine, histidine, glutamine and arginine) was analyzed by comparing plaque formation in the infected cells. The respective amino acids were additionally supplemented to a final concentration of 0.5 mM to Eagle's Basal Medium (EBM) without serum. The results were normalized to titers produced in EBM supplemented with all amino acids and set as 100%. The production of MV in this reference medium reached 4.6×10⁴ TCID₅₀ mL⁻¹ (Romano and Scarlata, 1973).

Three amino acids (arginine, methionine and glutamine) exerted major impact on the number of plaques and on virus production. The omission of these three amino acids always reduced MV yields, but the additional supplementation of arginine and glutamine to the higher concentrations of 0.5 mM, resulted in increased viral titers. In EBM without arginine, plaque formation was reduced to only 15%. An increase of arginine concentration up to 0.5 mM resulted in 2 fold higher plaque formation. The virus production in arginine-deprived medium resulted in a maximum titer of only 21 TCID₅₀ mL⁻¹ 48 h p.i. thus three log₁₀ units lower as compared to the reference. Methionine seems to be an essential amino acid for the production of MV in HeLa cells, as well. The absence of methionine resulted in early cell death, even before plaque formation became evident.

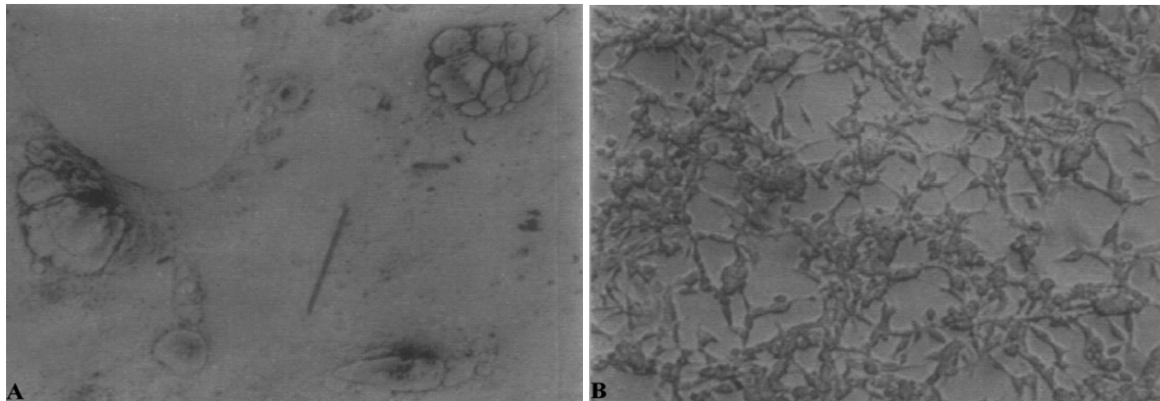


Fig. 4. Morphology of MV-infected Hep-2 cells cultured in different media (obtained from (Reissig *et al.*, 1956), with permission). Hep-2 cells grown in (A) Enders medium supplemented with bovine amniotic fluid, beef embryo extract, heated horse serum and Hanks balanced salt solution or (B) Eagles medium supplemented with amino acids and serum enrichments and calf serum. Cells were infected with the Edmonston strain of MV. Depicted are infected monolayer cultures ten days p.i. Phase contrast, about 60 x magnification.

For virus production without methionine, the titers did not exceed $32 \text{ TCID}_{50} \text{ mL}^{-1}$. The increase of methionine up to 0.5 mM did not increase maximum plaque formation above reference. However, addition of 0.5 mM methionine to cells with preceding methionine omission resulted in a three $\log_{10}$ units-fold increase of the virus titer (up to $3.2 \times 10^3 \text{ TCID}_{50} \text{ mL}^{-1}$) within the next 10 h. The third essential amino acid, glutamine, had similar effects as methionine. MV infection in the absence of glutamine was accompanied by early cell death without plaque formation. On the contrary, a glutamine concentration of 0.5 mM in the medium caused an increase of plaque formation to 1.5-fold higher numbers than the reference (Romano and Scarlata, 1973).

Other amino acids had no or a negative impact on MV replication and production. Tryptophan deprivation had no influence on plaque formation, while the addition of 0.5 mM of tryptophan was cytotoxic, resulting in rounded-up cells and a 50% decrease in plaque formation. The deprivation of isoleucine and threonine caused only 35% plaque formation and virus titers reached a maximum of 3.2×10^3 and $1 \times 10^3 \text{ TCID}_{50} \text{ mL}^{-1}$ 48 h p.i. respectively. The omission of the other amino acids resulted in 65-90% plaque formation, except for the deprivation of lysine, which revealed unchanged plaque formation. Increased concentrations of other amino acids had no influence on plaque formation. The highest MV titer ($1 \times 10^4 \text{ TCID}_{50} \text{ mL}^{-1}$), apart from the reference, was reached after 48 h p.i. in culture medium without tryptophan, lysine, cysteine and glutamine (Romano and Scarlata, 1973).

While reduced titers have been reported by the omission of serum, some components in serum seem to

be essential for MV production. As serum is complex and chemically undefined process instability and problems with batch to batch reproducibility are to be expected. Analyzing the available data, the amino acids arginine, methionine and glutamine seem to be important for high titers. However, this may vary with cell line and virus strain. In general, there are too little data available for robust interpretation. The special requirements considering amino acids of the host cells are still unknown, although glutamine has often been reported to be essential for MV production. Kinetic studies should be performed ideally with fully defined synthetic medium, which then enables implementation of optimal nutrient supply for MV production.

1.14. Impact of Multiplicity of Infection (MOI) on Measles Virus Production

The MOI describes the ratio of infectious agent (here: MV) to the infection target (cells). In general, MV infection for production purposes is performed at low MOI's around 0.02 (Kohn and Yassky, 1962; Brandriss *et al.*, 1982; Boriskin *et al.*, 1988; Sidorenko *et al.*, 1989; Trabelsi *et al.*, 2010).

Nakai *et al.* (1969) studied MV production by electron microscopy and varied the MOI. Bundles of filaments with a tubular nature, called tubules, were observed in infected cells' cytoplasm. These tubules were associated with the budding process. With ongoing MV replication, the tubules became larger and more numerous. Therefore, these tubules might be interpreted as MV nucleoprotein aggregates. Additionally, these tubules appeared in the nucleus, as well. These nuclear tubules were observed, when the cells were infected with

an MOI of 10. They appeared 24 h p.i. but they remained very sparse and poorly developed. The resulting extracellular virus titer was 10^4 pfu mL⁻¹ and no budding particles could be detected at any time. With lower MOI, the appearance of well-developed cytoplasmic tubules was observed 18 h p.i., extensive budding activity was evident after 30 h p.i. The titer of extracellular virus was three log₁₀ units higher (10^7 pfu mL⁻¹) than the one observed with an MOI of 10 (Nakai *et al.*, 1969).

Preferentially low MOI for producing high-titer MV was also demonstrated for BSC-1 cells. With an MOI of 1, the titers increased until day 2 p.i. and then remained stable. The highest values were about 1×10^6 and 2×10^5 pfu mL⁻¹ for cell-associated or extracellular virus, respectively. With a lower MOI of 0.01, the MV titers increased until day 3 and reached a maximum titer of 5×10^6 pfu cell⁻¹ for both cell-associated and extracellular virus (Nakai *et al.*, 1969).

For the production in Vero cell monolayers, the impact of MOI was analyzed by increasing the MOI in log₁₀ steps from 0.001-10 (Scott and Choppin, 1982). All maximum virus titers were between 10^7 and 10^8 pfu mL⁻¹. However, the highest MOI gave rise to the lowest maximum titer. In contrast, the best virus yield was produced with an MOI of 0.01, but reduction of the MOI elongated the time of virus production by about 5 days (Scott and Choppin, 1982).

In another study, the overall yield of MV production in Vero cell monolayers was higher with an MOI of 0.1 compared to an MOI of 10. However, using an MOI of 10, viral titers were higher until 48 h p.i. but production had to be terminated earlier (72 h MOI 10 vs. 96 h MOI 0.1) (Baczko and Lazzarini, 1979). In the same report, the titers in MA160 cell monolayers after infection with an MOI of 10 were always higher than titers reported after infection with MOI of 0.1. Again, using an MOI of 10, virus production had to be terminated one day earlier (96 h p.i. Vs 120 h p.i. respectively). In contrast, in MA160 suspension culture, the MV titer with MOI 10 were higher until 48 h p.i. while better titers appeared with an MOI of 0.1 starting 72 h p.i. until the end (Baczko and Lazzarini, 1979).

In MRC-5 cells the influence of the MOI was studied, too. The MOI was varied in steps of 0.01, 0.005 and 0.001. The highest titer was observed with an MOI of 0.01 (7.5×10^6 TCID₅₀ mL⁻¹), although the titers with an MOI of 0.005 and 0.001 were only slightly lower (1.8×10^6 and 7.5×10^5 TCID₅₀ mL⁻¹) (Trabelsi *et al.*, 2012).

In contrast to the results discussed above, with HeLa cells the more virus particles was produced the higher the MOI (0.01, 0.1 and 1) was. Interestingly, the maximum virus titer increased with increasing MOI to a more

drastic rate at 37°C (up to three log₁₀ units) than it did at 32.5°C (up to one log₁₀ unit) (Udem, 1984).

In most cases, a lower MOI was reported to be associated with higher viral titers. However, high MOI shortens production time, but increases the amount of virus needed for infection. Since economical consideration play a significant role in optimizing production processes, it should be noted that longer cultivation times and increased amounts for infection (higher MOI) cause higher production costs. On the other hand, with serial virus harvests as suggested above, the longer the time of production, the higher is the total generated virus amount. Thus, total yields with low MOI and serial virus harvests could be beneficial over longer production time in terms of production costs.

1.15. Impact of Culture Systems for Measles Virus Production

As already described for monolayer and suspension cultures, the choice of the culture system exerts considerable impact on virus production and the scalability of a process. Potential culture systems for MV production are in the range from small, simple and uncontrolled systems such as monolayer cultivation in T-flask or petri dishes up to highly controlled cultivation systems in large scales (e.g., Stirred Tank Reactors (STR)). Current MV vaccine production is performed mainly in adherent cells. As adherent cell lines usually need a growth surface to attach, the cultivation in a suspension culture such as in bioreactors can be supported by so called microcarriers. These microcarriers, on which the cells can grow, are small round or flat particles, which can easily be suspended by a stirrer.

1.16. Spinner Flask Cultivation

Boriskin *et al.* (1988) first reported MV production with cells cultivated on microcarriers. Quail embryo fibroblasts were cultivated on Cytolar-2 microcarriers and then infected with the MV strain Leningrad-16. Although the titers from roller bottle and microcarrier cultivation were within the same range, slightly higher virus titers were reached in roller cultures.

MV production with three different microcarriers (Cytodex 2, Micarcel G and Cytolar 2) had been investigated with quail embryo cells, as well (Sidorenko *et al.*, 1989). With Cytodex 2 carriers, the maximum virus titer (6×10^5 TCID₅₀ mL⁻¹) was reached on day 5 p.i.; afterwards, the virus titer decreased. With Micarcel G carrier, the highest maximum titer (8×10^5 TCID₅₀ mL⁻¹) was observed on day 4 p.i. For these microcarriers, the maximum MV titer remained constant for about 2 days and decreased following day 6 p.i. The lowest virus yield was measured on Cytolar 2 microcarriers.

Table 3. MV production performed with host cell cultivation on microcarrier. The respective bioreactor system, microcarrier and cell lined used are presented.

Culture system	Microcarrier	Cell line	MV strain	max. Titer ($\times 10^6$ TCID ₅₀ /mL)	Ref.
Spinner bottles	Cytolar-2	quail embryo cells	Leningrad 16	3.0	(Boriskin <i>et al.</i> , 1988)
	Cytodex 2	quail embryo cells	Leningrad 16	0.6	(Sidorenko <i>et al.</i> , 1989)
	Micarccl G	quail embryo cells	Leningrad 16	0.8	
	Cytolar 2	quail embryo cells	Leningrad 16	0.4	
Bioreactor (STR)	Cytodex 1	MRC-5	AIK-C	100,000.0	(Trabelsi <i>et al.</i> , 2010)
	Cytodex 1	MRC-5	AIK-C	40,000.0	(Trabelsi <i>et al.</i> , 2010)
	Cytodex 1	MRC-5	AIK-C	10.0	(Trabelsi <i>et al.</i> , 2012)
	Cytodex 1	Vero	Edmonston	50.0	(Mendonca <i>et al.</i> , 1994)

A maximum titer was reached at day 7 p.i. (4×10^5 TCID₅₀ mL⁻¹) and decreased thereafter. With microcarrier cultivation, a twofold enhancement of virus yields was obtained at best compared to roller cultures, reaching maximal production rates of 4.5-6.4 TCID₅₀ per cell. However, the virus production time and the harvest time was shortened with microcarrier systems. Furthermore, it was reported that MV yields could not be increased by higher microcarrier concentrations or cell densities. Sidorenko *et al.* (1989) further addressed the problem of determining the viable cell number after infection with MV. The cell number switched back and forth during MV production and determination of viable cell numbers by trypan blue exclusion was suggested to be not an adequate method. Since cells, which are stained blue, are supposed as dead, these cells could still release virus for an unknown time (Sidorenko *et al.*, 1989).

In another report, spinner flask cultures of MRC-5 cells were cultivated on Cytodex 1 microcarriers (Trabelsi *et al.*, 2010). In spinner flasks, the highest cell concentration was reached after 5 days of culture, which was chosen as time of infection. One day after infection with the AIK-C strain, the virus titer increased continuously until 6 days p.i. and reached a maximum titer of 1×10^{11} TCID₅₀ mL⁻¹ (Trabelsi *et al.*, 2010).

1.17. Bioreactor Cultivation

First data concerning MV production in a bioreactor were published in 1994 using an STR (Mendonca *et al.*, 1994). MV was amplified on MRC-5 cells in an STR grown on Cytodex 1 microcarriers (Trabelsi *et al.*, 2010), but also on a ready-to-use disposable fixed bed system (BelloCell) in Minimal Essential Medium (MEM) and 0.2% Bovine Serum Albumin (BSA). In the STR, the TCID₅₀ per cell and day was at minimum 15 times lower than in the BelloCell System (125 and 1,900 TCID₅₀ per cell and day, respectively). Shear stress was suggested to reduce the production yield in the STR (Trabelsi *et al.*, 2010).

Taken the available data together, spinner or bioreactor cultivations are sufficient systems for MV production. Although MV yields were not always higher compared to T-flaks or roller bottles, the bioreactor

system could still be advantageous considering the possibility of in-process control and direct regulation of e.g., pH and oxygenation. Additionally, serial harvesting could easily be automated. With further optimization of bioreactor cultivation, these systems seem to be very promising. MV yields produced with host cells cultivated on microcarriers in either spinner bottles or bioreactors are summarized in Table 3.

1.18. Impact of Variations in Culture Modes on Measles Virus Production

Controlled culture systems can work in different culture modes. This includes variation in temperature, stirrer speed, or different feeding and harvesting strategies, which all could influence virus stability and cell growth. The impact of the parameters temperature, shear force, feeding and serial harvests have already been investigated.

1.19. Impact of Culture Temperature

The various Edmonston derived vaccine strains have been further attenuated by additional passages in chicken embryo fibroblasts at a reduced temperature (32°C) (Katz, 2009). In some studies MV were also produced at lower temperature (Kohn, 1962; Boriskin *et al.*, 1988; Mendonca *et al.*, 1994; 2002), but MV production at 37°C is common as well (Black, 1959; Shishido *et al.*, 1967; Nakai *et al.*, 1969; Romano and Scarlata, 1973; Brandriss *et al.*, 1982). Consequently, the impact of the cultivation temperature during MV production has already been evaluated.

Amplification of attenuated and virulent Edmonston strains on BSC-1 cells has been reported to be similar at 34°C or 37°C (Rapp, 1964). A further comparative study of AIK-C strain MV production in MRC-5 cells at 34°C or 36°C in spinner cultures revealed slightly higher cell numbers with a cultivation temperature of 34°C, but MV titers were similar at both temperatures, again (Trabelsi *et al.*, 2010).

MV Hallé and Edmonston strains produced at 33°C or 37°C in Vero cell monolayers resulted in higher yields for the Hallé strain at 33°C. For the Edmonston strain, the yield was initially higher at

37°C, but the overall yield was better at 33°C, when a longer production phase was feasible (four instead of two days). For both virus strains, the cultivation at 37°C had to be terminated at least two days earlier than those at 33°C. This effect was supposed to be caused by the lytic nature of MV, which seems to be more intensive at higher temperatures (Scott and Choppin, 1982).

In HeLa cells MV yields were up to three folds higher (two log₁₀ units) at 32.5°C than at 37°C. Additionally, the author demonstrated that higher yields were not due to temperature sensitivity. The titers of 20 virus clones grown at 32.5°C were nearly the same (±10%) when plated at 32.5 and 37°C. Thus, it was suggested that host cell factors, such as protein synthesis, or a temperature-dependent host cell protease needed for the processing of virus polypeptides could be altered at the different temperatures (Udem, 1984).

1.20. Impact of Shear Forces

Mechanical shear forces have been published to increase MV yield from static monolayer cultures. Vigorous agitation could raise the virus titer about 2.5-12.5-fold in the supernatant. Since the mechanical forces did not increase cellular contaminants in the supernatant, it was suggested that the increase could be due to MV particles, whose budding was assisted by the help of the mechanical forces. As this vigorous agitation was performed in static cultures, higher virus yields in bioreactor cultivations with permanent shear forces could be suggested (Wechsler *et al.*, 1985). However, the agitation in static cultures was performed for the short time of harvest and not extensively during cultivation, thus it remains unclear, which effect a durable shear force during the whole cultivation period, as found in bioreactor systems, has got. Additionally, as serum is a protective agent concerning shear stress, it should be noted that the medium in the T-flasks had been replaced with serum-free medium 12-18 hours prior to the harvest (Wechsler *et al.*, 1985).

1.21. Impact of Culture Mode

Repeated batch and perfusion mode have been compared in an STR (Trabelsi *et al.*, 2010). These data revealed an approximately 6-fold higher virus yield in the repeated batch mode (125 TCID₅₀ per cell and day), compared to the perfusion mode (21 TCID₅₀ per cell and day).

In another study it has been demonstrated, that appropriate feeding of the cells is advantageous for MV production (Mendonca *et al.*, 2002). The analysis of the effects of glutamine or galactose feeding on the growth of Vero cells on Cytodex 1 microcarriers and subsequent MV production are summarized in **Table 4**

(Mendonca *et al.*, 2002). MV yields were successfully raised by increasing the cell number by feeding, as well. Infection on the fourth day of cultivation, when the batch cultivation was terminated, resulted in a virus titer of 5×10⁶ TCID₅₀ mL⁻¹ 5 days post infection. Without feeding, the cell number was drastically reduced until the 12th day of cultivation and infection of these cells resulted in a low virus yield 5 days p.i. However, with galactose and glutamine feeding, infection of 12 days old cultures resulted in a titer of 8×10⁶ TCID₅₀ mL⁻¹, again on day 5 p.i. The feeding resulted in a very significant reduction of the percentage of apoptotic cells on the day of infection. As illustrated in **Table 4**, the virus titer per cell increases with a reduced rate of apoptotic cells. These data reveal that an optimized production process for the cell line could lead to higher MV production rates (Mendonca *et al.*, 2002).

1.22. Impact of Harvesting Strategies

Overall virus yields have been reported to be enhanced by repeated harvest of released virus, when conditioned medium from infected monolayers has been replaced with fresh medium (Scott and Choppin, 1982). When considering the comparatively low half-life of the virus at 37°C (i.e., 2 h) (Black, 1959), this procedure might be promising. Interestingly, only slightly lower virus titers in the culture medium have been reported, when the medium was fully exchanged one hour prior to sampling (Black, 1959). This indicates some kind of equilibrium between release and decay and therefore suggests that a considerable amount of virus is wasted if serial harvesting from the culture medium is omitted. The effect of harvest time has also been investigated (Scott and Choppin, 1982). A single harvest on the fourth day p.i. resulted in yields of 28 pfu per cell for extracellular and 188 pfu per cell for cell-associated virus, which represents a ratio of released-to-cell-associated virus of about 1:6.7. Increasing the numbers of virus harvests from the supernatant resulted in higher released to cell-associated ratios for cumulative yields, respectively in values of 1:3.6 for one additional harvest on day 2 p.i. and 1:2.4 for three additional harvests on days 2.5, 3 and 3.5 p.i. With a total of 4 harvests, maximum numbers of 270 and 651 pfu per cell for extracellular and cell-associated virus, respectively, have been determined. This positive effect appeared always, when the time point of the first harvest was chosen at day 2 p.i. or later (Scott and Choppin, 1982). Daily harvesting of extracellular virus starting 24 h. p.i. resulted in reduced yields of 22 and 58 pfu cell⁻¹ for extracellular and cell-associated virus titers, respectively. The authors suggested that under these circumstances the medium removal one day after inoculation did not allow enough cells to become initially infected (Scott and Choppin, 1982).

Table 4. Effect of feeding on MV yield and cell viability, summary of data published by Mendonca *et al.* (2002). Presentation of the effect of feeding on apoptosis of cells and measles virus yield

Day of infection	Cell density at time of infection ($\times 10^5$ cells mL ⁻¹)	Apoptotic cells at day of infection (% of cells)	Feeding ^a	MV yield 5 days p.i.	
				($\times 10^6$ TCID ₅₀ mL ⁻¹)	(TCID ₅₀ per cell ^b)
4	5.1	2	no	5	9.8
12	3	88	no	0.00052	0.002
12	9.2	12	yes	8	8.7

^a; Fed with 4 mM galactose and 2 mM glutamine at days 4, 6, 8 and 10; ^b: Correlated to the cell number at time of infection

Table 5. Summary of published data for MV production. MV yields with respect to the MOI, the scale or mode of cultivation, virus strains and cell lines are presented. For better comparability, titers in pfu were converted in TCID₅₀ by multiplication with the factor 0.7

Max. Titer ($\times 10^6$ TCID ₅₀ /mL)	MOI	Cells/ MV strain	Cell-associated /extracellular MV	Scale/ mode of production	Remarks	Ref.
-	1	CV-1/ not specified	extracellular	150 cm ² T-flasks	(33°C p.i.), increased yield by vigorous agitation	(Wechsler <i>et al.</i> , 1985)
0.046	2-5	HeLa/ DP line of the Edmonston strain	-	16 cm rubber stoppered bottles, roller drum	examining amino acid deprivation during MV growth, (37°C p.i.)	(Romano and Scarlata, 1973)
0.36	0.01	quail embryo cells/ Leningrad-16 strain	-	siliconized 1 L spinner, working volume 0.5 L	Cytodex 2, Micarcel G, Cytolar 2 (each with 20 g/L)	(Sidorenko <i>et al.</i> , 1989)
0.56	0.025, 0.01	KB cells / Edmonston strain	cell-associated / extracellular	petri dishes (50 mm dish)	(37°C p.i.), MV inactivation kinetics at 37°C	(Kohn and Yassky, 1962)
0.63	1.5	Hep-2/ Edmonston strain	cell-associated / extracellular	-	growth curves of MV, characterization of temperature and pH-sensitivity, (37°C p.i.)	(Black, 1959)
3	1.4	Vero/ TYCSA strain (variant of the Toyoshima strain)	extracellular	roller tubes with 1 mL fluid	(37°C p.i.)	(Shishido <i>et al.</i> , 1967)
8	0,1	Vero / Edmonston strain	extracellular	T-flasks or spinner bottles at 60 rpm,	Cytodex 1 (2 g/L), cells are fed with galactose and glutamine, (33°C p.i.)	(Mendonca <i>et al.</i> , 2002)
8.6	0.02	U937 / Edmonston strain	cell-associated / extracellular, daily	suspension cells, spinner	(37°C p.i.)	(Brandriss <i>et al.</i> , 1982)
10	0.03	quail embryo cells / Leningrad-16 strain	-	stationary monolayer, roller or microcarrier cultures (0.5 L Techno cell stirrer bottles)	MC: Cytolar 2, (33°C p.i.)	(Boriskin <i>et al.</i> , 1988)
10	0.01, 0.001, 0.005	MRC-5/ AIK-C strain	-	STR 3 L working volume	Cytodex 1(3 g/L), MOI variation, different culture modes	(Trabelsi <i>et al.</i> , 2012)
14	0.01-1, 10	BSC-1, primary rhesus monkey kidney cells/ Edmonston	cell-associated / extracellular	4-ounce prescription bottles.	(37°C p.i.)	(Nakai <i>et al.</i> , 1969)
70	0.01-1	HeLa/Edmonston	cell-associated / extracellular	suspension cells	(37 and 32.5°C)	(Udem, 1984)
170	0.5	Vero/Edmonston strain	cell-associated / extracellular, until cytopathic effect (96-120 h p.i.)	Celligen cell culture 3.7 L, 60 rpm, for comparison also roux bottles'	Cytodex 1 (2 g/L), (33°C p.i.)	(Mendonca <i>et al.</i> , 1994)
230	0.1, 10	MA160, Vero/ Edmonston strain	cell-associated/ extracellular	monolayers (75 cm ²) and suspension cultures	-	(Baczko and Lazzarini, 1979)
400	0,001-10	HEp-2, AV3, BHK-21, PK-15, MDCK, Vero, BGIMK / Halle or Edmonston strain	mainly extracellular	flasks, monolayers	investigation of: FCS, temperature (33 and 37°C), buffer, insulin, virus strain, cell line, MOI	(Scott and Choppin, 1982)
100,000	0.01	MRC-5/ AIK-C strain	-	packed bed (BelloCell disposable); STR 2 L (working volume 1.2 L); spinner (working volume 0.5 L)	Cytodex 1(3 g/L), various cell culture modes, comparison of virus titers and cell concentrations, spinner: (36 and 34°C p.i.), bioreactor: (34°C p.i.)	(Trabelsi <i>et al.</i> , 2010)

However, this interpretation stays in contrast to reports describing that MV inactivation proceeds within few

hours and first progeny virus appears already 12 h after infection (Black, 1959).

The few available data for MV production suggest that feeding and repeated harvesting are advantageous for high virus yields. The culture temperature and shear stress can affect the total yield ambivalently. However, more detailed studies of these parameters might be considered.

1.23. Summary

Table 5 summarizes the currently available data for MV production with respect to the MOI, the scale or mode of cultivation, virus strains and cell lines. For better comparability, titers in pfu were converted in TCID₅₀ by multiplication with the factor 0.7.

2. CONCLUSION

Due to its ability of efficient tumor cell killing, MV has been identified as a potential oncolytic virus. Thus, there is considerable interest of a scalable, reproducible, optimized, standardized and automated production process for this promising agent.

Since being established for several decades, MV production for vaccination purposes is not exploiting the technical possibilities offered by modern cell culture technologies. However, simply up-scaling of the robust and established traditional technologies may be difficult when trying to push the viral yield by a factor of up to a million. In addition, potential second generation MV agents with modified tumor-selective cell-entry do most likely not infect traditional cell substrates used for producing parental vaccine MV.

Additionally, economic considerations play an important role for optimizing a biomedical production process. A great factor concerning costs is the scale of a process. It is a big difference whether production is performed in static systems as T-flasks and roller bottles, or in a bioreactor. For example, a bioreactor cultivation with 1 L working volume and Cytodex 1 microcarriers at a concentration of 3 g L⁻¹ confers the same surface area for cell growth as 44 T-flasks (each 300 cm² surface) or approx. 16 roller bottles (each 850 cm²). By this example, it is quite obvious that requirements for media, space and manpower are much less for the bioreactor cultivation. Bioreactor cultivation would be a good alternative to generate the amounts of MV needed for a routine therapeutic cancer treatment, since all other (including traditional) culture systems may be hard to scale up or even do not provide the necessary cell numbers and densities. The feasibility of bioreactor cultivation systems for MV production has already been demonstrated (Mendonca *et al.*, 1994; Mendonca *et al.*, 2002; Trabelsi *et al.*, 2010). As promising the bioreactor cultivation is, there is unfortunately only little data available referring to MV production in such systems. Additionally, further aspects like oxygen demand or shear stress have to be considered.

An optimization of different factors (temperature, MOI, addition of salts and repeated harvesting) has

already been investigated in the past. Some of these factors are very important, as e.g., the relative high sensitivity of the MV to temperatures (Black, 1959), or the potential to protect MV particles or promote infectivity by certain salts, respectively (Rapp *et al.*, 1965; Boriskin *et al.*, 1988). Referring to extracellular virus yields, the temperature sensitivity of the virus could be a critical parameter, as well. Virus in the supernatant is inactivated by elevated temperatures, which could be a reason for cell-associated virus yields being almost always reported to be higher than extracellular yields (Black, 1959; Rapp, 1964; Shishido *et al.*, 1967; Nakai *et al.*, 1969; Mendonca *et al.*, 1994). Since studies utilizing lowered temperatures during MV production revealed only slightly higher or similar virus titers (Rapp *et al.*, 1965; Scott and Choppin, 1982; Trabelsi *et al.*, 2010), this attempt may not be too promising. Having this in mind, it could be advantageous to integrate repeated or even continuous harvesting of extracellular virus with immediate cooling of the harvested virus into the process. Hence cell-associated virus can only be harvested once, total amounts of released virus from repeated harvests could exceed the amounts of cell-associated virus.

The addition of FCS provides serum proteins and essential amino acids for virus production. Accordingly, MV yields were almost always higher with serum supplemented medium (Scott and Choppin, 1982). Addition of serum is a critical factor, as it is of an unknown composition and therefore limits the reproducibility of a process. Furthermore, serum complicates the purification steps following production, since the product should be free of any contaminations when applied to human patients e.g., in cancer therapy. Thus, modern cultivation systems tend to rely on serum free cultivation, with complete serum free media being commercially available that can support comparable growth of host cells (Rourou *et al.*, 2007) and therefore high MV yields, too. Regarding medium composition, specific amino acids revealed a strong impact on MV growth (Romano and Scarlata, 1973) and appropriate feeding of Vero cells prior to MV infection also increased the MV yields (Mendonca *et al.*, 2002). However, the available data also indicate that a lot more factors influence MV production, accumulating in the need for detailed kinetic studies.

Besides exogenous factors the virus-cell system has to be optimized, as well. This includes that certain MV strains are adapted for growth in specific cell lines. Additionally, the ratio of virus per cell (MOI) is an important factor to consider. A low MOI around 0.01 reproducibly resulted in the highest MV titers (Nakai *et al.*, 1969; Baczko and Lazzarini, 1979; Scott and Choppin, 1982), but causes an elongated time of cultivation. Furthermore, high MOI reduced budding

of virus particles (Nakai *et al.*, 1969) and thus shifts the balance to cell-associated virus, which is more difficult to purify.

Processing for scale up systems in the field of cell culture has been performed over the past 50 years. Dynamic cultivations for mammalian cells have been operated from spinner systems up to bioreactors with working volumes up to several hundreds of liters (Eibl *et al.*, 2008). Development of microcarrier system made such cultivation modes possible for adherent cell cultures, as well. Moreover, the bioreactor cultivation should be an easy method to improve the space-time-yield and the microcarrier system seems to be a proper operation mode for the production of MV (Boriskin *et al.*, 1988; Sidorenko *et al.*, 1989; Mendonca *et al.*, 1994; Mendonca *et al.*, 2002; Trabelsi *et al.*, 2010). Much effort in standardized bioreactor cultivation with mammalian cells has been done. The growing use of disposable systems such as single-use bags for bioreactors, sensors, sampling or seed inoculums further advanced the cultivations (Eibl *et al.*, 2008).

For MV production, further efforts should concentrate on the following developments:

- A production process, which enables an easy scale up of the respective microcarrier systems
- As FCS is an undefined substance, MV should be produced in serum free medium
- Appropriate feeding of the cells during growth and virus production, which is based on detailed kinetic studies
- An established offline analytic would help integrating methods for online measurements and process automation
- The integration of an automated tool for serial or even continuous harvests

Solving these tasks should allow the set-up of an optimized, versatile production process, resulting in measles virus preparations intended for the use as oncolytic agents in the desired quality and quantities under affordable conditions.

2.1. Conflict of interest

The authors declare no conflict of interest.

3. ACKNOWLEDGEMENT

The researchers would like to thank the Hessen State Ministry of Higher Education, Research and the Arts for the financial support within the Hessen initiative for scientific and economic excellence (LOEWE-program).

4. REFERENCES

- Artenstein, A.W., 2009. Vaccines: A Biography. 1st Edn., Springer, New York, ISBN-10: 1441911073, pp: 401.
- Baczko, K. and R.A. Lazzarini, 1979. Efficient propagation of measles virus in suspension cultures. *J. Virol.*, 31: 854-855. PMID: 513197
- Black, F.L., 1959. Growth and stability of measles virus. *Virology*, 7: 184-192. DOI: 10.1016/0042-6822(59)90186-2
- Blechacz, B., P.L. Splinter, S. Greiner, R. Myers and K.W. Peng *et al.*, 2006. Engineered measles virus as a novel oncolytic viral therapy system for hepatocellular carcinoma. *Hepatology*, 44: 1465-1477. DOI: 10.1002/hep.21437
- Boriskin, Y.S., L.L. Steinberg, L.V. Dorofeeva, I.N. Zazorina and E.P. Barkova, 1988. Salt-induced enhancement of measles virus yields in cultured cells. *Arch Virol.*, 101: 131-136. DOI: 10.1007/BF01314658
- Brandriss, M.W., J.J. Schlesinger and S.E. Chapman, 1982. Growth of measles virus in a human macrophagelike cell line: U937. *Am. J. Pathol.*, 109: 179-183.
- Bronzino, J.D., 2000. The Biomedical Engineering Handbook. 2nd Edn., Springer, Boca Raton, ISBN-10: 354066808X, pp: 1408.
- Cathomen, T., B. Mrkic, D. Spohner, R. Drillien and R. Naef *et al.*, 1998. A matrix-less measles virus is infectious and elicits extensive cell fusion: Consequences for propagation in the brain. *EMBO J.*, 17: 3899-3908. DOI: 10.1093/emboj/17.14.3899
- Davis, J.J. and B. Fang, 2005. Oncolytic virotherapy for cancer treatment: challenges and solutions. *J. Gene Med.*, 7: 1380-1389. DOI: 10.1002/jgm.800
- Dorig, R.E., A. Marcil, A. Chopra and C.D. Richardson, 1993. The human CD46 molecule is a receptor for measles virus (Edmonston strain). *Cell*, 75: 295-305. DOI: 10.092-8674(93)80071-L
- Dubois-Dalcq, M. and T.S. Reese, 1975. Structural changes in the membrane of vero cells infected with a paramyxovirus. *J. Cell Biol.*, 67: 551-565. DOI: 10.1083/jcb.67.3.551
- Eagle, H., 1955. Nutrition needs of mammalian cells in tissue culture. *Science*, 122: 501-514. DOI: 10.1126/science.122.3168.501
- Eibl, R., D. Eibl and R. Pörtner, 2008. Cell and Tissue Reaction Engineering. 1st Edn., Springer, Berlin, ISBN-10: 3540681752, pp: 363.
- Enders, J.F. and Peebles, T.C., 1954. Propagation in tissue cultures of cytopathogenic agents from patients with measles. *Proc Soc Exp. Biol. Med.*, 86: 277-286. DOI: 10.3181/00379727-86-21073

- Enders, J.F., S.L. Katz, M.V. Milovanovic and A. Holloway, 1960. Studies on an attenuated measles-virus vaccine. I. Development and preparations of the vaccine: Technics for assay of effects of vaccination. *New England J. Med.*, 263: 153-159. DOI: 10.1056/NEJM196007282630401
- Gerlier, D., M.C. Trescol-Biemont, G. Varior-Krishnan, D. Nanche and I. Fugier-Vivier *et al.*, 1994. Efficient MHC class II-restricted presentation of measles virus to T cells relies on its targeting to its cellular receptor human CD46 and involves an endosomal pathway. *Cell Biol. Int.*, 18: 315-320. DOI: 10.1006/cbir.1994.1080
- Griffin, D.E. and M.B.A. Oldstone, 2008. *Measles: History and Basic Biology*. 1st Edn., Springer, Berlin Heidelberg, ISBN-10: 3540705228, pp: 200.
- Grote, D., S.J. Russell, T.I. Cornu, R. Cattaneo and R. Vile *et al.*, 2001. Live attenuated measles virus induces regression of human lymphoma xenografts in immunodeficient mice. *Blood*, 97: 3746-3754. DOI: 10.1182/blood.V97.12.3746
- Hasegawa, K., C. Hu, T. Nakamura, J.D. Marks and S.J. Russell *et al.*, 2007. Affinity thresholds for membrane fusion triggering by viral glycoproteins. *J. Virol.*, 81: 13149-13157. DOI: 10.1128/JVI.01415-07
- Kärber, G., 1931. Beitrag zur kollektiven Behandlung pharmakologischer Reihenversuche. *Arch. Exp. Path. Pharmacol.*, 162: 480-487. DOI: 10.1007/bf01863914
- Katz, S.L., 2009. John F. Enders and measles virus vaccine-a reminiscence. *Curr. Top Microbiol. Immunol.*, 329: 3-11. DOI: 10.1007/978-3-540-70523-9_4
- Katz, S.L., M.V. Milovanovic and J.F. Enders, 1958. Propagation of measles virus in cultures of chick embryo cells. *Proc. Soc. Exp. Biol. Med.*, 97: 23-29. DOI: 10.3181/00379727-97-23637
- Kohn, A. and D. Yassky, 1962. Growth of measles virus in KB cells. *Virology*, 17: 157-163. DOI: 10.1016/0042-6822(62)90092-2
- Kohn, A., 1962. Haemadsorption by measles syncytia. *Nature*, 193: 1088-1089. DOI: 10.1038/1931088a0
- Langfield, K.K., H.J. Walker, L.C. Gregory and M.J. Federspiel, 2011. Manufacture of measles viruses. *Methods Mol Biol*, 737: 345-366. DOI: 10.1007/978-1-61779-095-9_14
- Leber, M.F., S. Bossow, V.H. Leonard, K. Zaoui and C. Grossardt *et al.*, 2011. MicroRNA-sensitive oncolytic measles viruses for cancer-specific vector tropism. *Mol. Ther.*, 19: 1097-1106. PMID: 21468006
- Matumoto, M., 1966. Multiplication of measles virus in cell cultures. *Bacteriol Rev*, 30: 152-176.
- Mayer, E.D., 1960. Plaque formation by measles virus. *Virology*, 11: 634-638.
- Mendonca, R.Z., L.R. Vaz-de-Lima, M.I. Oliveira, C.A. Pereira and S. Hoshino-Shimizu, 1994. Studies on the efficiency of measles virus antigen production using VERO cell culture in a microcarrier system. *Braz J. Med. Biol. Res.*, 27: 1575-1587. PMID: 7874024
- Mendonca, R.Z., S.J. Arrozio, M.M. Antoniazzi, J.M.J. Ferreira and C.A. Pereira, 2002. Metabolic active-high density VERO cell cultures on microcarriers following apoptosis prevention by galactose/glutamine feeding. *J. Biotechnol.*, 97: 13-22. DOI: 10.1016/S0168-1656(02)00048-2
- Milovanovic, M.V., J.F. Enders and A. Mitus, 1957. Cultivation of measles virus in human amnion cells and in developing chick embryo. *Exp. Biol. Med.*, 95: 120-127. DOI: 10.3181/00379727-95-23140
- Morgan, E.M. and F. Rapp, 1977. Measles virus and its associated diseases. *Bacteriol. Rev.*, 41: 636-666.
- Msaouel, P., A. Dispenzieri and E. Galanis, 2009a. Clinical testing of engineered oncolytic measles virus strains in the treatment of cancer: an overview. *Curr Opin. Mol. Ther.*, 11: 43-53.
- Msaouel, P., I.D. Iankov, A. Dispenzieri and E. Galanis, 2011. Attenuated oncolytic measles virus strains as cancer therapeutics. *Curr. Pharm. Biotechnol.*, DOI: BSP/CPB/E-Pub/000272-13-1
- Msaouel, P., I.D. Iankov, C. Allen, J.C. Morris and von V. Messling *et al.*, 2009b. Engineered measles virus as a novel oncolytic therapy against prostate cancer. *Prostate*, 69: 82-91. DOI: 10.1002/pros.20857
- Mühlebach, M., M. Mateo, P. Sinn, S. Pruffer and K. Uhlig *et al.*, 2011. Adherens junction protein nectin-4 (PVRL4) is the epithelial receptor for measles virus. *Nature*, 480: 530-533. DOI: 10.1038/nature10639
- Mühlebach, M.D., T. Schaser, M. Zimmermann, S. Armeanu and K.M. Hanschmann *et al.*, 2010. Liver cancer protease activity profiles support therapeutic options with matrix metalloproteinase-activatable oncolytic measles virus. *Cancer Res.*, 70: 7620-7629. DOI: 10.1158/0008-5472.CAN-09-4650
- Nakai, T., F.L. Shand and A.F. Howatson, 1969. Development of measles virus *in vitro*. *Virology*, 38: 50-67. PMID: 4977468
- Nakamura, T., K.W. Peng, M. Harvey, S. Greiner and I.A. Lorimer *et al.*, 2005. Rescue and propagation of fully retargeted oncolytic measles viruses. *Nat Biotechnol.*, 23: 209-214. DOI: 10.1038/nbt1060
- Navaratnarajah, C.K., V.H. Leonard and R. Cattaneo, 2009. Measles virus glycoprotein complex assembly, receptor attachment, and cell entry. *Curr. Top Microbiol. Immunol.*, 329: 59-76. DOI: 10.1007/978-3-540-70523-9_4

- Nehring, D., P. Czermak, J. Vorlop and H. Lubben, 2004. Experimental study of a ceramic microsparging aeration system in a pilot-scale animal cell culture. *Biotechnol. Progress*, 20: 1710-1717. DOI: 10.1021/bp049762e
- Nehring, D., R. Gonzalez, R. Pörtner and P. Czermak, 2006. Experimental and modelling study of different process modes for retroviral production in a fixed bed reactor. *J. Biotechnol.*, 122: 239-253. DOI: 10.1016/j.jbiotec.2005.09.014
- Nehring, D., R. Pörtner, M. Schweizer, K. Cichutek and P. Czermak, 2009. Integrated in line filtration: A method to produce high concentrated retroviral vector titer supernatant. *Desalination*, 246: 241-247. DOI: 10.1016/j.desal.0000.00.000
- Noyce, R.S., D.G. Bondre, M.N. Ha, L.T. Lin and G. Sisson *et al.*, 2011. Tumor cell marker PVRL4 (Nectin 4) is an epithelial cell receptor for measles virus. *Plos. Pathog.*, 7: e1002240-e1002240. PMID: 21901103
- Peebles, T.C., K. McCarthy, J.F. Enders and A. Holloway, 1957. Behavior of monkeys after inoculation of virus derived from patients with measles and propagated in tissue culture together with observations on spontaneous infections of these animals by an agent exhibiting similar antigenic properties. *J. Immunol.*, 78: 63-74. PMID: 13406267
- Peng, K.W., C.J. TenEyck, E. Galanis, K.R. Kalli and L.C. Hartmann *et al.*, 2002. Intraperitoneal therapy of ovarian cancer using an engineered measles virus. *Cancer Res.*, 62: 4656-4662. PMID: 12183422
- Rapp, F., 1964. Plaque differentiation and replication of virulent and attenuated strains of measles virus. *J. Bacteriol.*, 88: 1448-1458.
- Rapp, F., J.S. Butel and C. Wallis, 1965. Protection of measles virus by sulfate ions against thermal inactivation. *J. Bacteriol.*, 90: 132-135.
- Reed, L.J. and H. Muench, 1938. A simple method of estimating fifty percent endpoint dilution. *Am. J. Epidemiol.*, 27: 493-497.
- Reissig, M., F.L. Black and J.L. Melnick, 1956. Formation of multinucleated giant cells in measles virus infected cultures deprived of glutamine. *Virology*, 2: 836-838. PMID: 13392528
- Romano, N. and G. Scarlata, 1973. Amino acids requirements of measles virus in HeLa cells. *Arch Gesamte Virusforsch*, 43: 359-366. DOI: 10.1007/BF01556153
- Rourou, S., A.V.D. Ark, T.V.D. Velden and H. Kallel, 2007. A microcarrier cell culture process for propagating rabies virus in Vero cells grown in a stirred bioreactor under fully animal component free conditions. *Vaccine*, 25: 3879-3889. DOI: 10.1016/j.vaccine.2007.01.086
- Russell, S.J., A.K. Fielding, K.W. Peng and D. Grote, 2010. Method for limiting the growth of cancer cells using an attenuated measles virus. Mayo Foundation for Medical Education and Research.
- Sabella, C., 2010. Measles: Not just a childhood rash. *Cleve Clin. J. Med.*, 77: 207-213. PMID: 20200172 DOI: 10.3949/ccjm.77a.09123
- Scheld, W.M., R.J. Whitley and C.M. Marra, 2004. *Infections of the Central Nervous System*. 3rd Edn., Lippincott Williams and Wilkins, Philadelphia, ISBN-10: 0781743273, pp: 939.
- Scott, J.V. and P.W. Choppin, 1982. Enhanced yields of measles virus from cultured cells. *J. Virol. Methods*, 5: 173-179. DOI: 0166-0934(82)90007-6
- Shishido, A., K. Yamanouchi, M. Hikita, T. Sato, A. Fukuda and F. Kobune, 1967. Development of a cell culture system susceptible to measles, canine distemper and rinderpest viruses. *Arch. Virol.*, 22: 364-380. DOI: 10.1007/BF01242957
- Sidorenko, E.S., L.V. Dorofeeva, T.I. Kaptsova, L.L. Steinberg and I.N. Zazorina *et al.*, 1989. Experimental-scale measles and mumps vaccine production on microcarrier-grown cells. *Vaccine*, 7: 554-556. DOI: 10.1016/0264-410X(89)90282-X
- Springfield, C., V. von Messling, M. Frenzke, G. Ungerechts and C.J. Buchholz *et al.*, 2006. Oncolytic efficacy and enhanced safety of measles virus activated by tumor-secreted matrix metalloproteinases. *Cancer Res.*, 66: 7694-7700. DOI: 10.1158/0008-5472.CAN-06-0538
- Tatsuo, H., N. Ono, K. Tanaka and Y. Yanagi, 2000. SLAM (CDw150) is a cellular receptor for measles virus. *Nature*, 406: 893-897. DOI: 10.1038/35022579
- Trabelsi, K., S. Majoul, F. Charfi and H. Kallel, 2010. Measles virus production in MRC-5 cells grown on microcarrier in a stirred bioreactor. *ESACT Proc.*, 4: 765-769. DOI: 10.1007/978-90-481-3419-9_133
- Trabelsi, K., S. Majoul, S. Rourou and H. Kallel, 2012. Development of a measles vaccine production process in MRC-5 cells grown on Cytodex1 microcarriers and in a stirred bioreactor. *Applied Microbiol. Biotechnol.*, 93: 1031-1040. DOI: 10.1007/s00253-011-3574-y
- Udem, S.A., 1984. Measles virus: Conditions for the propagation and purification of infectious virus in high yield. *J. Virol. Methods*, 8: 123-136. DOI: 10.1016/0166-0934(84)90046-6
- Wechsler, S.L., D.M. Lambert, M.S. Galinski, B.E. Heineke and A.L. Lambert *et al.*, 1985. A Simple method for increased recovery of purified paramyxovirus virions. *J. Virol. Methods*, 12: 179-182. DOI: 10.1016/0166-0934(85)90018-7